Efficient Glycosidation of a Phenyl Thiosialoside Donor with Diphenyl Sulfoxide and Triflic Anhydride in Dichloromethane

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David Crich* and Wenju Li

Department of Chemistry, University of Illinois at Chicago, 845 West Taylor Street, Chicago, Illinois 60607-7061

dcrich@uic.edu

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ABSTRACT



The formation of sialic acid glycosides with a thiosialic acid derivative, diphenyl sulfoxide, and trifluoromethanesulfonic anhydride is reported. With an excess of diphenyl sulfoxide, glycal formation can be completely suppressed and excellent yields are obtained for coupling to a wide range of primary, secondary, and tertiary acceptors.

The sialic acids, discovered in vertebrates by Blix and Klenk in 1941, are a family of more than 40 2-keto-3-deoxynonulosonic acids that are incorporated in oligosaccharides and glycoconjugates which play important biological roles in high animals and human beings.1 The lead member of the series, N-acetylneuraminic acid (Neu5Ac), is often found at the terminal positions of glycoproteins and glycolipids and connected to galactosides or 2-acetamido-galactosides by α - $(2\rightarrow 3)$ or $\alpha(2\rightarrow 6)$ linkages. The biological significance of the sialic acid glycosides combines with the widely acknowledged difficulty inherent in their synthesis to render this class of glycosidic bonds one of the most widely studied in the field of carbohydrate chemistry.^{2,3} Both the challenge and the importance of the problem are readily appreciated from the application of almost every major class of glycosidic bond forming reaction to the sialic acid glycosides in recent years.²⁻⁴ Thioglycosides are no exception to this rule, with most common promoters having been evaluated in this respect,^{2–4} with the exceptions of the recent 1-benzenesulfinyl piperidine (BSP)/triflic anhydride system developed in this laboratory,^{5,6} the related diphenyl sulfoxide (Ph₂SO)/ triflic anhydride method as adapted to the thioglycosides by van Boom,^{6,7} and the Kahne sulfoxide method.^{6,8} We are interested in developing rapid, low-temperature α -selective sialylations from thioglycoside donors, avoiding the use of acetonitrile and its stereodirecting effect, which is common to much work in the area,^{2,3,4,9} and report here on our preliminary observations.

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Among the various 2-thiosialyl donors, we chose methyl 2-β-phenylthio-4,7,8,9-tetra-O-acetyl N,N-diacetylneuraminate (1), because of its crystalline nature and enhanced selectivity, compared to the N-monoacetyl counterpart.^{3n,10} Initially, we investigated our BSP/Tf₂O/TTBP activation method,⁷ but saw no activation even after changing the hydroxy protecting groups from esters to more armed silyl ethers. We explored therefore the more potent promotion system Ph₂O/Tf₂O, which was developed by Gin for dehydrative glycosylation strategies,¹¹ including sialytations.^{11d} Hoping to generate the β -anomeric sially triflate as intermediate species, we first applied a stoichiometric quantity of the Ph₂SO/Tf₂O combination in the presence of the mild, hindered base 2,4,6-tri-tert-butylpyrimidine (TTBP).¹² Unfortunately, while activation was complete at -78 °C in a matter of minutes, it was followed by immediate and quantitative elimination to the glycal 3. However, raising the amount of Ph₂SO to 3 equiv, with otherwise unchanged conditions, we found that the coupling reaction proceeded in excellent yield and good selectivity on subsequent addition of 2-propanol (Table 1, entry 1, 97%, 2.3:1 α/β). We postulate that the initial intermediate oxacarbenium ion/ anomeric triflate pair 2 undergoes instantaneous decomposition to the glycal **3** in the absence of a suitable nucleophile. However, in the presence of excess sulfoxide this species is trapped to give two covalent sulfonium salts 4 and 5, which serve as resevoirs for the oxacarbenium ion in the subsequent glycosylation. Low-temperature NMR experiments in CD₂-Cl₂ support this hypothesis, with two apparently isomeric species being observed in the ratio 1.5:1 when the reaction was conducted with 3 equiv of Ph₂SO.¹³ These intermediates,

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Table 1.	Coupling the Presence of	300% Diphenyl Sulfoxide
entry	acceptor	product: % yield ^a (α:β) ^ь
1	OH	19: 97% (2.3:1)
2	6 MeOH 7	20: 94% (1.5:1)
3		21: 91% (1:1)
4	но-СД	22: 89% (3.8:1)
5		1 ₁₇ 23: 92% (2:1)
6	Bno Bno OMe	24: 97% (1.2:1)
7		25: 92% (6:1)
8		26: 94% (10:1)
9		27: 92% (onlyα)°
10	HO OBN HO BNO OME	28: 76% (1:1.2) ^d
11	HO O HO O OMe	29: 81% (1:1.7) ^d
12		30: 82% (1:7)
13	BnO OBn HO BnO OMe 18	-

^{*a*} Isolated yields. ^{*b*} Determined by ¹H NMR on the crude reaction mixture. ^{*c*} Coupled to the 6-OH. ^{*d*} Coupled to the 3-OH.

tentatively assigned to 4 and 5, were immediately converted to the two methyl glycosides, also in the ratio 1.5:1, on addition of methanol at -78 °C.

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This protocol,¹⁴ employing 3 molar equiv of diphenyl sulfoxide, was then applied to a range of acceptors (Table 1), with anomeric stereochemistry assigned on the basis of the chemical shift of the sialic acid H-3,15 H-4,16 and, preferably, the sialic acid ${}^{3}J_{C1,H3ax}$.¹⁷ With the exception of alcohol 18, for which no coupled product was isolated, the isolated yields of these coupling reactions ranged from good to excellent. Stereoselectivity, on the other hand, varied considerably and was very substrate dependent. Optimal α -selectivities were obtained for the important galactose 6-OH (Table 1, entries 7-9) but, interestingly, considerably lower selectivity was obtained with a glucose 6-OH derivative (Table 1, entry 6). With the 3,4-diols no improvement in selectivity was seen on switching from the 2,6-di-O-benzyl ether 15 to the 2,6-di-O-propargyl ether, despite the reduced steric bulk of the latter acceptor (Table 1, entries 10 and 11).¹⁸ Perhaps not surprisingly, the worst selectivity was obtained with the less reactive 4-OH group (Table 1, entry 12). All in all, it is clear that correct stereochemical matching¹⁹ of the acceptor with the donor is important in these couplings, as is becoming increasingly apparent in glycosylation reactions in general.²⁰

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Та	able 2. The Effect of Additives and Sulfoxides i) activator (3 equiv), TTBP (2 equiv), AcO OAc CO_2Me Tf_2O (1.1 equiv) AcO OAc					
_	ii) isc	propanol (2 equiv)		19		
_	entry	activator	solvent (temp.)	% yield ^a $(\alpha:\beta)^{b}$		
-	1	O I Ph ^S Ph	CH ₂ Cl ₂ (- 60 °C)	82% (2.2:1)		
	2	O Bh S Ph	CH ₂ Cl ₂ /CH ₃ CN 1:1 (- 78 °C)	89% (1.8:1)		
	3	O Bh ^S -Ph	CH ₃ CH ₂ CN (- 78 °C)	77% (2:1)		
	4	(4-NO2-Ph)PhSO	CH ₂ Cl ₂ (- 78 °C)	75% (2.7:1)		
	5	(4-MeO-Ph)PhSO	CH ₂ Cl ₂ (- 78 °C)	50% (2:1)		
	6		CH ₂ Cl ₂ (- 78 °C)	50% (6:1)		

^a Isolated yields. ^b Determined by ¹H NMR on the crude reaction mixture.

While a marginally better yield was obtained when the standard solvent was replaced by a 1:1 mixture of acetonitrile and dichloromethane, the $\alpha:\beta$ selectivity was diminished on coupling to 2-propanol (Table 2, entries 1 and 2). The use of propionitrile alone as solvent also failed to bring about the desired improvement in selectivity (Table 2, entry 3). Reverting to dichloromethane as solvent, the replacement of diphenyl sulfoxide by the more electron-deficient 4-nitrophenyl phenyl sulfoxide brought about a small increase in selectivity (Table 2, entry 4), while the more electronrich 4-methoxyphenyl phenyl sulfoxide gave no increase in selectivity and a substantially lower yield (Table 2, entry 5). The use of dibenzothiopene oxide, while giving only a 50% yield of coupled product under the standard activation conditions, did result, however, in a significant increase in selectivity to 6:1 in favor of the α -glycoside with model alcohol 2-propanol (Table 2, entry 6). The improvement of selectivity on replacing diphenyl sulfoxide by dibenzothiophene oxide must result from a differing population and/or reactivity of the glycosyl sulfonium ions corresponding to 4 and 5. The relatively low yield obtained with dibenzothiophene oxide as activating agent resulted from poor conversion of the donor 1, which is presumably due to reduced electrophilicity of sulfoxide O-trifluoromethane-

⁽¹³⁾ The H-3eq peaks of the two anomers (at δ 3.1 and 2.9) are characteristic of the two species in the ¹H NMR spectrum.

⁽¹⁴⁾ General experimental procedure for glycosylation reactions: The 2-thiosialic acid donor 1 (0.11 mmol), diphenyl sulfoxide (Ph₂SO, 0.32 mmol), TTBP (0.22 mmol), and activated 4 Å powdered molecular sieves were mixed in anhydrous dichloromethane (2 mL) and cooled to -78 °C under positive argon pressure. The mixture was stirred at -78 °C for 30 min before Tf₂O (0.12 mmol) was added. After 10 min, a solution of acceptor (0.22 mmol) in anhydrous dichloromethane (1 mL) was added. The reaction mixture was stirred from 1 to 6 h at -78 °C and then warmed to room temperature, filtered, and washed with saturated aqueous NaHCO₃ solution and brine. The organic layer was dried over Na₂SO₄ and concentrated under reduced pressure. The glycosides were isolated by silica gel column chromatography or by preparative HPLC.

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^a Isolated yields. ^b Determined by ¹H NMR on the crude reaction mixture.

sulfonylated adduct. Accordingly, we developed a protocol in which donor **1** was activated with Tf_2O in the presence of both diphenyl sulfoxide and dibenzothiophene oxide and applied to several coupling reactions (Table 3).

Unfortunately, the improvement in selectivity seen with 2-propanol did not extend to any of the other examples investigated.

In summary, we have shown that the challenging α -sialylation with 2-thiosialic acid donors can be efficiently performed by using the diphenyl sulfoxide/trifluoromethanesulfonic anhydride promotion system. The use of excess diphenyl sulfoxide is shown to be important in these couplings and serves to suppress the formation of the glycal by trapping the first-formed oxacarbenium ion, as suggested by low-temperature NMR studies. Sialylations of a series of alcohol nucleophiles showed satisfactory yields and variable anomeric selectivities that are sensitive to the exact nature of the acceptors. With this new procedure, the Neu5Ac $\alpha(2\rightarrow 6)$ Gal glycosidic linkages can be installed with excellent yield and selectivity. Sialylation of secondary sugar acceptors and even the simple tertiary alcohol 1-adamantanol proceeded with good yield but modest selectivity.

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Note Added after ASAP Publication. There were symbols missing in the heading and body of Tables 1 and 3 in the version published ASAP February 2, 2006; the corrected version was published ASAP February 6, 2006.

Supporting Information Available: Full experimental details and characterization data. This material is available free of charge via the Internet at http://pubs.acs.org.

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